

graphed over Si gel plates eluted with  $\text{CHCl}_3$  as eluent. After repeated crystallization from MeOH, **3** (100 mg) was obtained, mp 226–229°;  $[\alpha]_D^{23} + 41^\circ$  (c 0.86;  $\text{CHCl}_3$ ); IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3300 (hydroxyl), 1750 ( $\gamma$ -lactone), 1675 (cyclopentenone), 1632, 1618 (olefinic bonds), 1418, 1390, 1325, 1290, 1231, 1205, 1100, 1085, 1045, 1010, 865, 765, 684. UV  $\lambda_{\text{max}}^{\text{EtOH}}$  nm (log  $\epsilon$ ): 257 (4.18) (cyclopentenone chromophore), 235 (4.02) ( $\alpha,\beta$ -unsaturated lactone).  $^1\text{H}$  NMR (90 MHz,  $\text{CDCl}_3$ ): see Table 1. EIMS (direct inlet) 75 eV,  $m/z$  (rel. int.): 260 ( $\text{M}^+$ , 100), 245 (6), 242 (13), 227 (8), 214 (8), 199 (23), 197 (19), 186 (14), 185 (13), 171 (14), 143 (11), 135 (66), 126 (60), 107 (40), 105 (16), 98 (16), 91 (65), 77 (28), 69 (81), 65 (30), 51 (30), 44 (39). (Found: C, 68.94; H, 6.30.  $\text{C}_{15}\text{H}_{16}\text{O}_4$  requires: C, 69.21; H, 6.20%.)

**Acknowledgements**—We thank Dr J. Borja, Department of Botany, Faculty of Pharmacy (Madrid) for the identification of plant materials and Miss M. D. Casado and Miss M. Plaza for recording the  $^1\text{H}$  NMR spectra.

#### REFERENCES

1. Pinar, M., Rico, M. and Rodríguez, B. (1982) *Phytochemistry* **21**, 735.
2. Bagirov, V. Y., Sheichenko, V. I., Casanova, R. Y. and Pimenov, M. G. (1978) *Khim. Prir. Soedin.* 811.
3. Serkerov, S. and Sheichenko, V. I. (1970) *Khim. Prir. Soedin.* 425.
4. Bagirov, V. Y., Sheichenko, V. I., Abdullaeva, I. K. and Pimenov, M. G. (1980) *Khim. Prir. Soedin.* 834.

*Phytochemistry*, Vol. 21, No. 7, pp. 1804–1806, 1982.  
Printed in Great Britain.

0031-9422/82/071804-03\$03.00/0  
© 1982 Pergamon Press Ltd.

## ISOLATION AND STRUCTURE OF STEPHALIC ACID, A NEW CLERODANE DITERPENE FROM *STEVIA POLYCEPHALA*\*

ENRIQUE ANGELES, KIRSTEN FOLTING,† PAUL A. GRIECO,† JOHN C. HUFFMAN,† R. MIRANDA and MANUEL SALMÓN‡

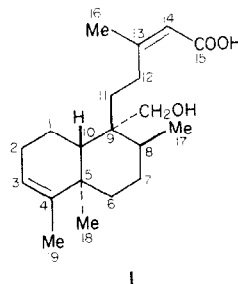
Escuela Nacional de Estudios Profesionales Cuautitlan, UNAM Cuautitlan Izcalli, Campo 1, Departamento de Química, Estado de México, México; †Department of Chemistry and Molecular Structure Center, Indiana University, Bloomington, IN 47405, U.S.A.; ‡Instituto de Química, Universidad Nacional Autónoma de México, México 20, D. F., México.

(Revised received 9 October 1981)

**Key Word Index**—*Stevia polycephala*; Compositae; stephalic acid; clerodane derivatives; diterpenes.

**Abstract**—From the methanolic extract of *Stevia polycephala* a new clerodane-type diterpene, stephalic acid, was isolated. The structure and stereochemistry were determined by a combination of spectral data and single-crystal X-ray analysis.

The *Stevia* genus is one of the largest found in Mexico [1]. Sesquiterpene lactones [2–4] and diterpenes [5] have been isolated from the non-polar or chloroform fractions of Mexican *Stevia* species. Isohumelene and new  $\alpha$ -longipinene derivatives have been isolated from *Stevia polycephala* [6]. In our continuing search for new natural products from *Stevia* species, we examined the polar fraction of the methanolic extract of *S. polycephala* collected in the state of Tlaxcala, Mexico, and have isolated a new diterpene, stephalic acid (**1**), which possesses the clerodane carbon skeleton.



Fractionation of the methanol extract of *S. polycephala* with ethyl acetate and chromatographic separation employing Si gel provided a new diter-

\*Contribution No. 563.

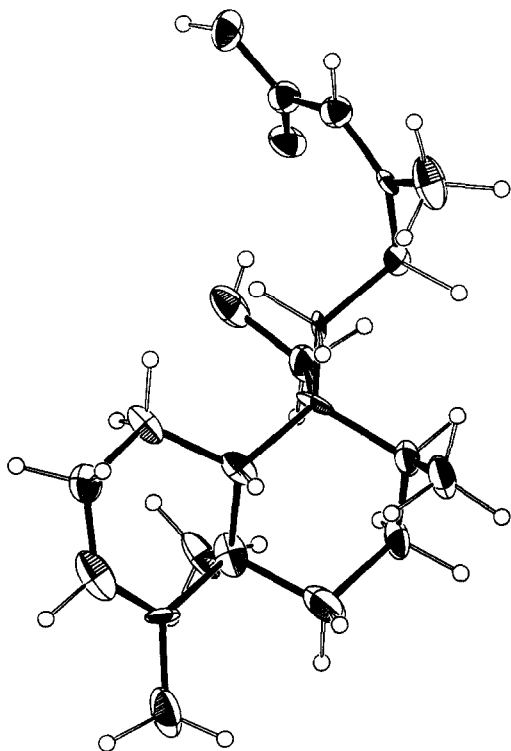
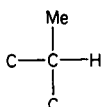


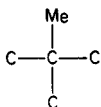
Fig. 1. ORTEP view of stephalic acid.

pene. Stephalic acid (1),  $C_{20}H_{32}O_3$ ,  $M^+$  at  $m/z$  320, was obtained as a crystalline solid, mp  $183.5$ – $185.0^\circ$ ,  $[\alpha]_D^{25} + 25.8$  (c 1.0,  $CHCl_3$ ). The IR spectrum ( $CDCl_3$ ) of 1 exhibited a three-proton singlet at  $\delta$  1.93 and a one-proton singlet at 1.73 which together with IR ( $CHCl_3$ ) bands at 1615, 1685, and  $2300$ – $3300\text{ cm}^{-1}$  and a UV (MeOH) absorption at  $\lambda_{max}$  202 nm ( $\epsilon$  9600) suggested the presence of a  $\beta$ -alkyl,  $\beta$ -methyl acrylic acid unit.

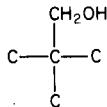
The presence of a two-proton AB quartet centered at  $\delta$  4.00 in the  $^1H$  NMR spectrum, together with an absorption band at  $3420\text{ cm}^{-1}$  in the IR spectrum, indicated the presence of a hydroxymethyl group attached to a quaternary carbon. Further examination of the  $^1H$  NMR spectrum revealed the presence of a secondary methyl group ( $\delta$  1.59), a methyl group ( $\delta$  1.14) attached to a quaternary carbon, and an olefinic methyl group (1.59) of which the olefin was part of a six-membered ring. The above data led to five partial structures A–E.



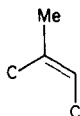
A



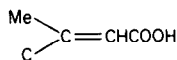
B



C



D



E

Unambiguous structural assignment was obtained by carrying out a single-crystal X-ray analysis of 1 (Fig. 1). Stephalic acid crystallizes in the monoclinic space group  $P2_1$ , with  $a = 8.51$  (2),  $b = 7.52$  (2),  $c = 14.65$  (4) Å,  $b = 100.86$  (11),  $V = 920.5$  Å<sup>3</sup>, giving  $D_{calc} = 1.156\text{ g/cm}^3$  for  $Z = 2$ . All measurements were at  $-165^\circ$ . A total of 2290 unique intensities were collected using a  $\theta$ - $2\theta$  continuous scan technique with the following values: scan rate =  $2.5/\text{min}$ , 10-sec stationary scans at extremes of scan,  $2.0^\circ$  scan width. All data were collected for  $+h$ ,  $+k$ ,  $\pm l$  in the range  $6 \leq 20 \leq 45$  using Mo  $K_\alpha$  radiation ( $\lambda = 0.71069$  Å). The structure was solved by direct methods (MULTAN) and refined by full-matrix techniques. All atoms were located and refined, with the thermal parameters of the hydrogen atoms fixed. Final residuals were  $R(F) = 0.077$  and  $R_w(F) = 0.059$  [7]. No attempt was made to assign the absolute configuration.

#### EXPERIMENTAL

**Isolation of stephalic acid.** *Stevia polycephala* Bertol was collected in the state of Tlaxcala, Mexico, during Sept. 1977. The dried whole plant (2 kg) was extracted with 15.0 l. of warm MeOH. The extract was filtered, concentrated to 2.0 l., and washed in a liquid-liquid extractor with 1.0 l. of hexane which was discarded. The MeOH extract was evaporated to dryness under red. pres. providing 110 g of a syrupy green oil. The part soluble in warm EtOAc (80 g) was chromatographed on 1.0 kg of Si gel (packed in EtOAc- $C_6H_6$ , 1:9). The column was successively eluted, taking fractions of ca. 500 ml: fractions 1–5 (EtOAc- $C_6H_6$ , 1:9), fractions 6–10 (EtOAc- $C_6H_6$ , 3:7), fractions 11–20 (EtOAc- $C_6H_6$ , 6:4), fractions 21–26 (EtOAc), fractions 27–37 (EtOAc-MeOH, 4:1).

Fractions 18–25 which exhibited one major spot on TLC analysis were combined and rechromatographed on Si gel. The EtOAc- $C_6H_6$  (3:2) fractions were combined and the crude material obtained upon evaporation of the solvent was submitted to prep. TLC on Si gel ( $C_6H_6$ -EtOAc, 4:1, three developments). This produced 40 mg of a gum which upon crystallization from EtOAc-hexane, provided pure stephalic acid, mp  $183.5$ – $185.0^\circ$ :  $[\alpha]_D^{25} + 217.0^\circ$  (c 1.0,  $CHCl_3$ ); UV  $\lambda_{max}^{MeOH}$  nm: 202 ( $\epsilon$  9600); IR  $\nu_{max}^{CHCl_3}$   $cm^{-1}$ : 3420, 3300–2300, 1685, 1615;  $^1H$  NMR ( $CDCl_3$ ):  $\delta$  1.93 (s, 3H, H-17), 1.14 (s, 3H, H-18), 1.59 (br s, 3H, H-19), 1.93 (s, 3H, H-16), 3.23 (br t, 1H,  $J = 10\text{ Hz}$ ), 4.00 (ABq, 2H,  $J = 12\text{ Hz}$ ,  $\Delta\nu_{AB} = 12.5\text{ Hz}$ ,  $\nu_{AB} = 3.20$  for s, 3.33, 3.53 (s, 1H, H-14); MS  $m/z$  (rel. int.): 320 [ $M^+$ ] (5), 302 [ $M - H_2O$ ] (18), 284 [ $M - 2H_2O$ ] (15), 202 (14), 191 (13), 184 (35), 161 (14), 147 (22), 133 (23), 121 (34), 120 (30), 119 (35), 109 (28), 107 (48), 100 (31), 95 (100), 93 (36), 91 (28), 80 (42), 69 (26), 67 (25), 55 (40), 41 (41). (Found: C, 74.81; H, 9.96.  $C_{20}H_{32}O_3$  requires: C, 74.96; H, 10.06%.)

Complete X-ray crystallographic data are available in microfiche form from the Indiana University Chemistry Library. Request M.S.C. Report 8040 when ordering.

**Acknowledgement**—This investigation was supported by a Public Health Service Research Grant from the National Cancer Institute.

#### REFERENCES

- Matuda, E. (1958) *Bol. Soc. Bot. Mex.* **23**, 55.
- Salmón, M., Diaz, E. and Ortega, A. (1973) *J. Org. Chem.* **38**, 1759.

3. Salmón, M., Ortega, A. and Diaz, A. (1975) *Rev. Latinoam. Quim.* **6**, 45.
4. Salmón, M., Diaz, E. and Ortega, A. (1977) *Rev. Latinoam. Quim.* **8**, 172.
5. Salmon, M., Angeles, E., Ortega, A. and García de la Mora, G. (unpublished, observation).
6. Bohlmann, F., Suwita, A., Natsu, A. A., Czerson, H. and Suwita, A. (1977) *Chem. Ber.* **110**, 3572.
7. Huffman, J. C., Lewis, L. N. and Caulton, K. G. (1980) *Inorg. Chem.* **19**, 2755.

*Phytochemistry*, Vol. 21, No. 7, pp. 1806–1807, 1982.  
Printed in Great Britain.

0031-9422/82/071806-02\$03.00/0  
© 1982 Pergamon Press Ltd.

## A NEW SECO-LABDANE DERIVATIVE FROM *ATHRIXIA ELATA*\*

FERDINAND BOHLMANN, MICHAEL WALLMEYER and JASMIN JAKUPOVIC

Institute for Organic Chemistry, Technical University of Berlin, D-1000 Berlin 12, W. Germany

(Received 20 October 1981)

**Key Word Index**—*Athrixia elata*; Compositae; diterpenes; seco-labdane derivative; triterpenes; 2 $\beta$ -hydroxyerythrodil.

**Abstract**—*Athrixia elata* afforded, in addition to known compounds, a new seco-labdane derivative and a new triterpene.

From the mainly South African genus *Athrixia* (tribe Inuleae, subtribe Athrixiinae) some species have been investigated chemically [1]. In addition to triterpenes and thymol derivatives some diterpenes related to kaurene were present. We have now studied the constituents of *A. elata* Sond. The roots afforded the thymol derivatives 1–3, friedelin, dammadienone and the corresponding acetate, *ent*-kaurenic acid (6) and the 9, 11-dehydro derivative 7 and 4-formylath-

rixinone (8) [1]. The aerial parts gave squalene, germacrene D, caryophyllene,  $\alpha$ -humulene, the cinnamates 4 and 5 [2], the triterpene 9 and the seco-labdane derivative 10. The structure of 9 followed from the molecular formula and the <sup>1</sup>H NMR spectrum (Table 1), which was close to that of erythrodil [3]. The position and the stereochemistry of the additional hydroxyl group followed from the couplings of H-2 and from double resonance experiments. Ac-

Table 1. <sup>1</sup>H NMR spectral data of compound 9 (400 MHz, CDCl<sub>3</sub>, TMS as int. standard)

H-1	2.07 dd	H-23'	3.56 d
H-1'	2.14 dd	H-24	1.01 s
H-2	4.09 ddd	H-25	1.03 s
H-3	3.23 d	H-26	0.96 s
H-9	1.49 ddd	H-27	1.16 s
H-11	1.87 ddd	H-28	1.26 s
H-11'	1.99 ddd	H-29	0.87 s
H-12	5.21 dd	H-30	0.89 s
H-23	3.21 d		

*J* (Hz): 1, 1' = 12.5; 1', 2 = 2.5; 2, 3 = 5; 9, 11 = 6.5; 9, 11' = 12; 11, 11' = 17; 11, 12 = 11', 12 = 3; 12, 12' = 11.

Table 2. <sup>1</sup>H NMR spectral data of compound 10 (400 MHz, TMS as int. standard)

	CDCl <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>
H-5	2.03 t	2.06 t
H-6	2.43 d	2.33 d
H-9	2.66 dd	2.63 dd
H-11	2.39 ddd	2.46 ddd
H-11'	2.61 ddd	2.75 ddd
H-12	5.23 dd(br)	5.51 dd(br)
H-14	6.30 dd	6.51 dd
H-15	4.94 d	5.01 d
H-15'	5.09 d	5.17 d
H-16	1.72 s(br)	1.84 s(br)
H-17	2.12 s	1.90 s
H-18	0.98 s	0.74 s
H-19	0.99 s	0.73 s
H-20	0.95 s	0.92 s

*J* (Hz): 5, 6 = 4.5; 9, 11 = 2; 9, 11' = 11.5; 11, 11' = 13; 11, 12 = 5; 11', 12 = 8.5; 14, 15 = 11; 14, 15' = 17.

\*Part 435 in the series "Naturally Occurring Terpene Derivatives". For Part 434 see Bohlmann, F. and Gupta, R. K. (1982) *Phytochemistry* **21** (in press).